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Key Points:

- Remobilization of microbes is dependent on both their antecedent accumulation in streambed sediments and the magnitude of stream flow
- Storm events mobilize microbes from streambed stores that are replenished during the falling limb of the storm hydrograph and baseflow
- Our particle tracking mobile-immobile model captures exchanges between the surface water, the hyporheic zone and deeper streambed

Supporting Information:

Supporting Information may be found in the online version of this article.

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Modeling Contaminant Microbes in Rivers During Both Baseflow and Stormflow

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Abstract Rivers transport contaminant microorganisms (including fecal indicator bacteria and human pathogens) long distances downstream of diffuse and point sources, posing a human health risk. We present a mobile-immobile model that incorporates transport as well as immobilization and remobilization of contaminant microbes and other fine particles during baseflow and stormflow. During baseflow conditions, hyporheic exchange flow causes particles to accumulate in streambed sediments. Remobilization of stored particles from streambed sediments occurs slowly during baseflow via hyporheic exchange flow, while remobilization is vastly increased during stormflow. Model predictions are compared to observations over a range of artificial and natural flood events in the dairy contaminated Topehaehae Stream, New Zealand. The model outputs closely matched timing and magnitude of *E. coli* and turbidity observations through multiple high-flow events. By accounting for both state-of-flow and hyporheic exchange processes, the model provides a valuable framework for predicting particle and contaminant microbe behavior in streams.

Plain Language Summary Contaminant microorganisms, including the bacterial indicator *E. coli*, and various disease-causing bacteria, viruses, and pathogens, are highly episodic in rivers—with typically low-contaminant microorganism concentrations during low flows that are 100—fold or more increased during storms. At low flow, microbes and other fine particles tend to accumulate steadily in near-surface streambed sediments (the "hyporheic zone"), but these stores are remobilized by accelerating currents as flow increases. We developed a numerical model framework to represent exchanges of particles and microbes between water and the streambed sediments under variable states of flow—including the deeper streambed as well as the hyporheic zone. Our model was able to capture microbial behavior measured over both a natural storm event and a series of three artificial floods (without any wash-in from land) in the dairy-contaminated Topehaehae Stream, New Zealand. Our modeling approach provides a useful framework for predicting microbial behavior and associated hazards within rivers and downstream waters.

1. Introduction

Public health risks from the presence of contaminant microorganisms in waters, such as human pathogenic bacteria, parasites, or viruses, are a global concern (Ramirez-Castillo et al., 2015). Although rivers can transport microorganisms to long distances, timescales of retention and persistence in streambed sediments prior to downstream transport can range from days to years (Haggerty et al., 2002; Jamieson et al., 2004; Petersen & Hubbart, 2020), extending potential risks to long timescales after initial contamination of the stream. Stormflow events are known to resuspend retained microbes (Davies-Colley et al., 2008; McKergow & Davies-Colley, 2010) with the movement of microbes hypothesized to be linked to bed-mobilizing flows that remobilize sediments and attached microbes (Cho et al., 2010; de Brauwere et al., 2014; Zhou et al., 2017). However, microbes are also remobilized during steady-state baseflow (i.e., subcritical flow conditions) below the bed-mobilizing threshold (Bradshaw et al., 2016; Fluke et al., 2019; Muirhead & Meenken, 2018; Park et al., 2017), therefore providing evidence of other co-occurring processes that lead to measurable concentrations of microbes in streams during baseflow. Hence, appropriately characterizing transport and retention of contaminant microbes during both baseflow and stormflow conditions is required for predicting in-stream contamination and assessing microbial hazards.

Hyporheic exchange flow, the transport of solutes, and fine particles, including microbes, to and from the water column via flowpaths through streambed sediments (Boano et al., 2014; Haggerty et al., 2002; Krause





et al., 2011, 2017), is an important process, often not considered in models of contaminant microbe behavior in streams. For example, J. D. Drummond et al. (2018) demonstrated that hyporheic exchange flow can cause up to 66% of contaminant microbe inputs into an agriculturally impacted stream to persist for years under baseflow conditions. In fact, the hyporheic zone is an important ecotone for a diverse set of processes that provide oppositions. tunities for the self-purification of rivers, including the storage and degradation of pollutants and modulation of metabolic stream processes (Lewandowski et al., 2019). Turbulence near to the surface water-sediment interface and advective transport pathways caused by pressure variations at the streambed surface are the main reasons for the exchange of microbes between surface water and streambed sediments and other transient storage area (Roche et al., 2019) although there are a wide range of hydrostatic and hydrodynamic forces considered as hypor

§ heic exchange processes (Boano et al., 2014; Grant et al., 2011). However, current models used to predict water quality in freshwaters normally assume that microbes can only be transported into streambed sediments by incog poration into aggregates that settle by gravity (e.g., see review by Cho et al., 2016). Hyporheic exchange processed can furthermore result in baseflow remobilization of microbes (J. D. Drummond et al., 2015, 2018) althoug models that incorporate both baseflow and stormflow attribute baseflow remobilization to other processes, suc as biofilm sloughing (Kim et al., 2017; Park et al., 2017).

Available modeling frameworks do not account for baseflow and stormflow fine particle transport, including contaminant microbes, that simulate hyporheic exchange, immobilization, and remobilization processes. A suits able model should be parsimonious, that is, use as few parameters as possible to characterize the key processes and match the data so as to narrow the available parameter space and provide confidence in the best-fit value € (J. Drummond et al., 2019; Kelleher et al., 2019). An appropriate model framework needs to address not only fine particle mobilization and transport during storm events, but also differing transport mechanisms between the rising and falling limb of a storm hydrograph. As it is not yet possible to measure the transport of particles at this level of detail during a storm event, there is scope for model-based assessments to describe transport behavior of particles over events. During a storm event, particles retained within the hyporheic zone are partially remobilized (J. D. Drummond et al., 2017; Filoso et al., 2015; Harvey et al., 2012; Larsen et al., 2015). During the rising lim stormflow hydrograph, there is net remobilization of retained contaminant microbes (Lamba et al., 2015). Build

stormflow hydrograph, there is net remobilization of retained contaminant microbes (Lamba et al., 2015). Building on this field evidence, we hypothesize that although deposition into the hyporheic zone takes place during the rising limb of the storm event, deposited particles will follow advective porewater paths back into the water column instead of transporting deeper into the streambed. Moreover, we hypothesize that on the falling limb of transporting deeper into the streambed. Moreover, we hypothesize that on the falling limb of transporting deeper into the streambed. Moreover, we hypothesize that on the falling limb of transport into the deeper streambed, and remobilization back to the water column are all taking place simultanely ously. Finally, we aim to explore here how baseflow remobilization occurs not merely after the critical thresholds for mobilizing streambed sediments is exceeded, but also because of hyporheic exchange processes combined with the increased remobilization observed during a storm event.

To test the above hypotheses, we developed and validated a particle tracking mobile-immobile model for in-streamy conditions. This new model framework builds on the mobile-immobile approach (Haggerty & Gorelick, 1995) and Genuchten & Wierenga, 1976) and incorporates hyporheic exchange processes to simulate particles in the surface water, the hyporheic region, and the deeper streambed. We aim to capture both the sharp rising limb and slower falling limb of contaminant microbes over storm hydrographs and test our hypotheses on the controlling mechanisms of microbial transport under baseflow and stormflow within a single model framework. We apply this model to in-stream E. coli and turbidity data for a dairy-contaminated stream in response to (a) a triplet engineered flood pulses at 1-day intervals and (b) a two-peaked natural stormflow event. We demonstrate the engineered flood pulses at 1-day intervals and (b) a two-peaked natural stormflow event. We demonstrate the engineered flood pulses at 1-day

DRUMMOND ET AL. 2 of 10

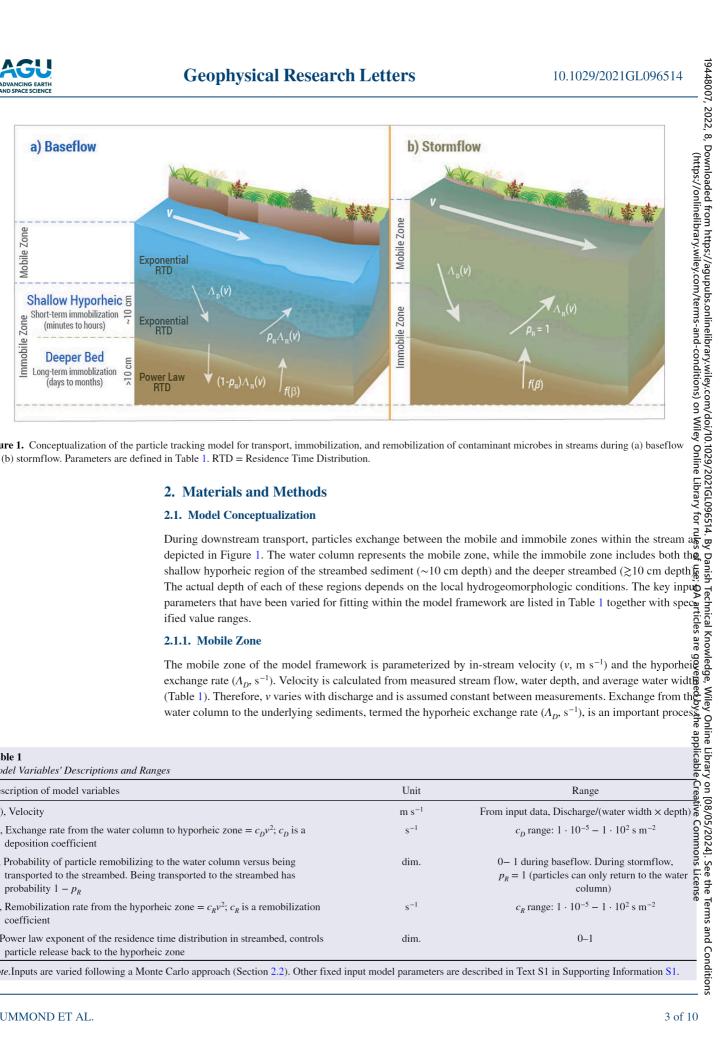


Figure 1. Conceptualization of the particle tracking model for transport, immobilization, and remobilization of contaminant microbes in streams during (a) baseflow and (b) stormflow. Parameters are defined in Table 1. RTD = Residence Time Distribution.

Table	1	
Model	Variables' Descriptions and Ranges	

Description of model variables	Unit	Range
v(t), Velocity	m s ⁻¹	From input data, Discharge/(water width × depth)
Λ_D . Exchange rate from the water column to hyporheic zone = $c_D v^2; c_D$ is a deposition coefficient	s^{-1}	c_D range: $1 \cdot 10^{-5} - 1 \cdot 10^2$ s m ⁻²
p_R . Probability of particle remobilizing to the water column versus being transported to the streambed. Being transported to the streambed has probability $1-p_R$	dim.	0-1 during baseflow. During stormflow, $p_R = 1$ (particles can only return to the water column)
$\Lambda_R,$ Remobilization rate from the hyporheic zone = $c_R v^2; c_R$ is a remobilization coefficient	s^{-1}	c_R range: $1 \cdot 10^{-5} - 1 \cdot 10^2$ s m ⁻²
β , Power law exponent of the residence time distribution in streambed, controls particle release back to the hyporheic zone	dim.	0–1

Note. Inputs are varied following a Monte Carlo approach (Section 2.2). Other fixed input model parameters are described in Text S1 in Supporting Information S1.

DRUMMOND ET AL. 3 of 10



that leads to the deposition of microbes and other fine particles with low settling velocities (Boano et al., 2014; $\frac{N}{N}$, $\frac{N}{N}$). D. Drummond et al., 2020). Residence times in the water column are exponentially distributed with an average exchange rate into the hyporheic zone proportional to the square of in-stream velocity (Text S1 in Supporting Information S1, Arnon et al., 2013; Packman et al., 2004), calculated as $\Lambda_D = c_D v^2$, where c_D is a deposition coefficient (Table 1).

2.1.2. Immobile Zone

Contaminant microbes transported into the shallow hyporheic region can either transport further into the deeper streambed or return to the water column, controlled by a resuspension probability p_R that can range from 0 to 1 (Table 1). A p_R of 1 signifies that particles can only follow the transport path back to the water column and conversely, a p_R of 0 signifies that particles can only transport into the deeper streambed. Residence times in the hyporheic zone are exponentially distributed with an average exchange rate back to the water column or into the deeper streambed proportional to the square of in-stream velocity (termed the remobilization rate, Λ_R), based on previous observations of fine sediment remobilization from the streambed (Arnon et al., 2013; Cardena's contact and 1995: Cho et al., 2010). The remobilization rate is calculated as $\Lambda_R = c_R v^2$, where c_R is the remobilization of the square of the streambed (Arnon et al., 2013; Cardena's contact and the square of the square et al., 1995; Cho et al., 2010). The remobilization rate is calculated as $\Lambda_R = c_R v^2$, where c_R is the remobilization coefficient. Here, we do not require that a critical threshold is met before microbes can be remobilized from the hyporheic zone to either the deeper streambed or water column. This lack of a critical threshold is supported by previous laboratory and fieldwork that demonstrate the remobilization of fine particles during baseflow (Brad[®] shaw et al., 2016; J. D. Drummond et al., 2015; Fluke et al., 2019; Muirhead & Meenken, 2018; Park et al., 2017

The deeper streambed is characterized by a power law residence time distribution (RTD), based on field obser vations of microbial retention and release from streambed sediments (Aquino et al., 2015; J. D. Drummon& Aubeneau, & Packman, 2014; J. D. Drummond, Davies-Colley, et al., 2014; Haggerty et al., 2002), and compare to an exponential distribution that allows for a wider range of times when contaminant microbes are release back to the hyporheic zone. As soon as microbes are released from the deeper streambed to the hyporheic zone they will again be subject to transport to the water column or back to the streambed with a probability p_R and remobilization rate, Λ_{P}

2.1.3. Stormflow

During stormflow, the same transport processes were considered in the model, but we ran three different scenare ios to test our hypotheses on how transport of microbes may differ between the rising and falling limbs of the storm hydrograph. We first assessed model outputs without any changes from baseflow parameters (scenaria 1), and then only allowed deposited particles in the hyporheic zone to transport back to the water column by setting $p_R = 1$ (Section 2.1.2, Figure 1b) during both the rising and falling limbs (scenario 2) and only the rising limb (scenario 3) of the storm hydrograph. This adjustment forces retained or deposited microbes already in the hyporheic zone to remobilize back to the water column instead of deeper into the streambed, aligning with field observations (J. D. Drummond et al., 2015, 2017; Filoso et al., 2015; Harvey et al., 2012; Lamba et al., 2015).

2.2. In-Stream Field Studies of Contaminant Microbe Transport Dynamics

Following the fitting procedure outlined in J. Drummond et al., 2019, we performed several simulations (Texts 2) in Supporting Information S1) with parameter sets constrained to match the in-stream measurements of the coli and turbidity during artificial floods (Section 2.2.1) and a natural storm event (Section 2.2.2) in a dair cow-impacted stream in New Zealand. The three scenarios for stormflow as described in 2.1.3 were evaluated for the artificial floods and natural storm event E. coli data, separately. Then, the best-fit scenario was used to five formation of the artificial floods.

An experiment with artificial flood pulses was conducted in the Topehaehae Stream (median flow $\sim 2.6 \cdot 10^{2}$ long the first assertion of the artificial flood pulses was conducted in the Topehaehae Stream (median flow $\sim 2.6 \cdot 10^{2}$ long the first assertion of the artificial flood pulses was conducted in the Topehaehae Stream (median flow $\sim 2.6 \cdot 10^{2}$ long the first assertion of the artificial flood pulses were conducted on 3 successive days by opening a release (Muirhea During stormflow, the same transport processes were considered in the model, but we ran three different scenars

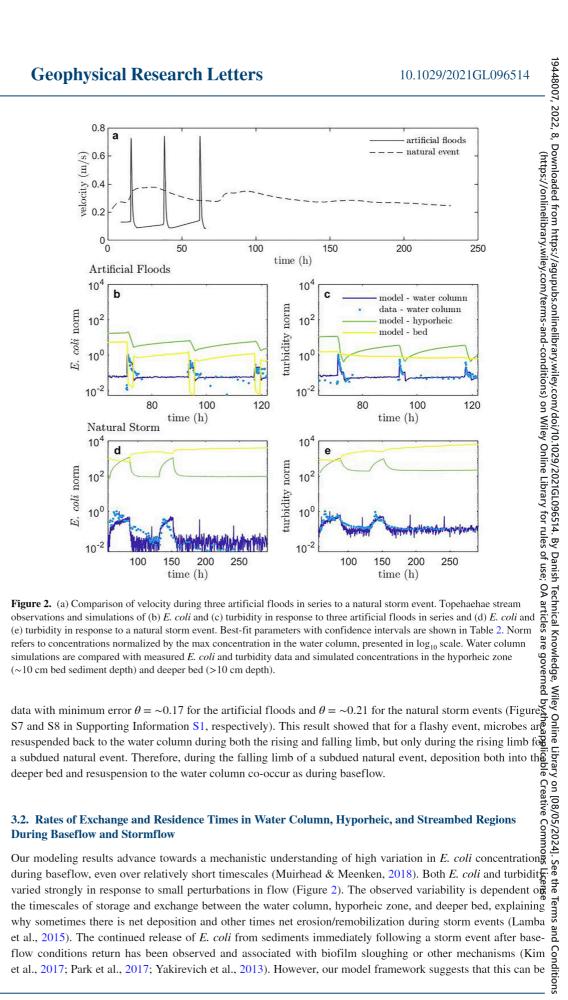
(Muirhead et al., 2004). The artificial flood pulses were conducted on 3 successive days by opening a release valve over 30 min, keeping it open for 20 min, and closing it over 10 min. Flow increased ~5-6- fold from $7.7 \cdot 10^2$ to $4.3 \cdot 10^3$ L s⁻¹ during each pulse. The water level, turbidity, and *E. coli* were measured at several sites downstream, and we focus on the furthest site 2.5 km downstream from the reservoir. The average stream width of the study reach was 5.8 m. The increase in the water level during the flood event was confined within the channel

DRUMMOND ET AL. 4 of 10

Table 2 Best-Fit Parameters, Defined in Table 1, and Associated Confidence Intervals Calculated as ± the Standard Deviation of the Best 0.05% Fits for Water Column E. coli and Turbidity Measurements During Three Artificial Flood Events in Series in Topehaehae Stream

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Table 2 Best-Fit Parameters, Defined in Table	2 1, and Associated Confi	idence Intervals Calculated	as ± the Standard Deviation	n of the Best 0.05% Fits fo	r Water Column E.	
oli and Turbidity Measurements During Three Artificial Flood Events in Series in Topehaehae Stream						
Parameter		Artificial floods E. coli Turbidity		Natural storm event E. coli Turbidit		
Best-fit model parameters		L. con	Turbiarty	E. con	Turolarty	
$c_D ext{ (s m}^{-2})$		$1.7 \cdot 10^{-1} \pm 4.5 \cdot 10^{-2}$	$1.3 \cdot 10^{-1} \pm 5.1 \cdot 10^{-2}$	$8.0 \cdot 10^1 \pm 3.1 \cdot 10^1$	$1.6 \cdot 10^1 \pm 2.6 \cdot 10^1$	
$c_R ext{ (s m}^{-2})$		$3.7 \cdot 10^{-3} + 1.2 \cdot 10^{-3}$	$6.7 \cdot 10^{-3} + 2.0 \cdot 10^{-3}$	$1.1 \cdot 10^{-2} + 1.6 \cdot 10^{-1}$	$3.0 \cdot 10^{-3} + 4.5 \cdot 10^{-3}$	
p_R		$1.1 \cdot 10^{-2} \pm 8.0 \cdot 10^{-3}$	$1.2 \cdot 10^{-2} \pm 3.6 \cdot 10^{-3}$	$4.3 \cdot 10^{-1} \pm 1.1 \cdot 10^{-1}$	$6.5 \cdot 10^{-1} \pm 1.6 \cdot 10^{-1}$	
β		$9.9 \cdot 10^{-1} \pm 1.9 \cdot 10^{-1}$	$6.3 \cdot 10^{-1} \pm 1.4 \cdot 10^{-1}$	$2.4 \cdot 10^{-1} \pm 1.2 \cdot 10^{-1}$	$1.6 \cdot 10^{-1} \pm 5.6 \cdot 10^{-2}$	
Temporally averaged rates and residen	nce time estimates					
Λ_D baseflow (s ⁻¹)		$2.9 \cdot 10^{-3}$	$2.2 \cdot 10^{-3}$	$3.9 \cdot 10^{0}$	$7.6 \cdot 10^{-1}$	
Λ_D peak stormflow (s ⁻¹)		$9.3 \cdot 10^{-2}$	$7.0 \cdot 10^{-2}$	$1.1 \cdot 10^{1}$	$2.3 \cdot 10^{0}$	
Λ_R baseflow (s ⁻¹)		$6.3 \cdot 10^{-5}$	$1.1 \cdot 10^{-4}$	$5.3 \cdot 10^{-4}$	$1.5 \cdot 10^{-4}$	
Λ_R peak stormflow (s ⁻¹)		$2.0 \cdot 10^{-3}$	$3.7 \cdot 10^{-3}$	$1.6 \cdot 10^{-3}$	$4.3 \cdot 10^{-4}$	
Water column residence time basef	low $(1/\Lambda_D, \text{hour})$	$1.0 \cdot 10^{-1}$	$1.3 \cdot 10^{-1}$	$7.1 \cdot 10^{-5}$	$3.7 \cdot 10^{-4}$	
Water column residence time storm	flow $(1/\Lambda_D, \text{hour})$	$3.0 \cdot 10^{-3}$	$4.0 \cdot 10^{-3}$	$2.4 \cdot 10^{-5}$	$1.2 \cdot 10^{-4}$	
Hyporheic residence time baseflow	$(1/\Lambda_R, \text{hour})$	$4.4 \cdot 10^{0}$	$2.5 \cdot 10^{0}$	$5.2 \cdot 10^{-1}$	$1.9 \cdot 10^{0}$	
Hyporheic residence time peak stor	CI (1/A 1)	1.4.10-1	7 6 10-2	4 = 40 1	C 4 10-1	
	tes were calculated base without any overb	d on average best-fit model pank flow, thereby allowing	parameters.	1.7-10 ⁻¹	e particles and E. co	
	without any overb	d on average best-fit model pank flow, thereby allowing burces. For more experiments	parameters. ng for a focused study of ental details, see Text S3	n remobilization of fine	e particles and E. coation S1.	
Note. Rates and residence time estima	without any overb from in-channel so	d on average best-fit model pank flow, thereby allowing burces. For more experimental comments of the comments	parameters. ng for a focused study of ental details, see Text S3	n remobilization of fine	e particles and E. co	
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	were analyzed for to this precipitation $5.0 \cdot 10^3 \mathrm{L s^{-1}}$ at the turbidity during the width lower slope	E. coli and turbidity by ton event, stream flow rose first flood peak and 3.5 e natural storm event we and silty-sand bed as a	the same methods as for use 10-fold from a basel \cdot 10 ³ L s ⁻¹ at the second are made downstream an ecompared to the artificial	the artificial flood exp flow of 5.0 · 10 ² L s ⁻ peak. In-stream measured in a reach with shallond flood sampling site.	eriments. In respon- before the event rements of <i>E. coli</i> are ower depth, increase Therefore, microbi	
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DRUMMOND ET AL. 5 of 10



flow conditions return has been observed and associated with biofilm sloughing or other mechanisms (Kim et al., 2017; Park et al., 2017; Yakirevich et al., 2013). However, our model framework suggests that this can be

DRUMMOND ET AL. 6 of 10



more simply explained by hyporheic exchange, not only into, but also out of, the streambed, depending on flow conditions.

The model matched the experimentally observed decrease in peak concentration with each subsequent artificial flood pulse, representing the depletion of *E. coli* and fine sediment (i.e., turbidity) from the streambed sed of ments (Figures 2b and 2c, respectively). In general, best-fit parameters for the artificial floods were very similar for *E. coli* and turbidity (Table 2). Overall, all model parameters fell within the expected ranges. Specifically, advective exchange rates were 2.9 · 10⁻³ and 2.2 · 10⁻³ s⁻¹ for *E. coli* and turbidity, respectively (Table 2) advective exchange of water and turbulence at the surface water-sediment bed interface controls the transport operation of the streambed of appropriately characterizing this important process. Residence in the model framework in that it is capable of appropriately characterizing this important process. Residence in the model framework in that it is capable of appropriately characterizing this important process. Residence times in the hyporheic zone actually increase with the increased stream flow velocity during the flood release but with lower retention times in the hyporheic region before being released back to the water column (Table 2). During baseflow, microbes and fine sediments were mainly transported from the hyporheic zone into the streambed and not immediately back to the water column as shown by a very low p_R of ~1 · 10⁻² for both *E. coli* and tribidity (Table 2). A low p_R aligns with previous observations of microbial transport during baseflow, using a power law RTD (e.g., J. D. Drummond, Aubeneau, & Packman, 2014; J. D. Drummond, Davies-College, et al., 2014). Moreover, the remobilization rate (A_R) was lower than the deposition rate into the hyporheic zone (Boano et al., 2014; E. 2014). D. Drummond, Aubeneau, & Packman, 2014; J. D. Drummond, Davies-Colley, et al., 2014). Therefore, during a streambed,

baseflow, both E. coli and fine sediments transport into the hyporheic region and within hours also into the deepe streambed, where retention times are longer and release back into the hyporheic zone is slow and can take hours to months (J. D. Drummond et al., 2018; J. Drummond et al., 2019). One difference between the measured microbes

months (J. D. Drummond et al., 2018; J. Drummond et al., 2019). One difference between the measured microbes and fine sediments (turbidity) was a slightly lower power law slope in the deeper streambed, β, for E. coli that turbidity, suggesting increased retention and slower release of E. coli back to the hyporheic zone. A lower for E. coli can either be explained by the increased attachment of microbes that excrete extracellular polymeries substances, which could decrease their release from the deeper streambed to the hyporheic region and eventually back to the water column (Battin et al., 2016; Eboigbodin & Biggs, 2008) or alternatively, a result of the longer term inactivation of E. coli in the streambed.

Higher E. coli and turbidity values in the hyporheic and deeper streambed regions were obtained in simulation between the three than in the water column (Figures 2b–2e), matching field observations. However, the interplay between the three model regions differs between E. coli and turbidity (Figures 2b and 2c, respectively) even with small difference model regions differs between E. coli and turbidity (Figures 2b and 2c, respectively) even with small difference in parameter values. Overall, we observed that unsurprisingly, the hyporheic zone is much more dynamic than the more stable deeper bed, exhibiting sharper changes in concentration during the flood events. The new model framework matches the artificial flood data and demonstrates how the hyporheic zone connects the surface water of the more stable deeper bed and regulates the slow release of contaminant microbes back into the water column during baseflow and fast release during stormflow, appropriately representing the transport and accumulation behaviore of both microbes and fine particles.

Our work supports the concept that remobilization of E. coli from the sediment bed during natural storm events of both microbes and fine particles.

only leads to partial removal as has been observed experimentally (J. D. Drummond, Aubeneau, & Packman, 2014 J. D. Drummond, Davies-Colley, et al., 2014; Stocker et al., 2018). We were able to provide some insight into microbial release during a natural storm event and deposition co-occurring with remobilization during the falling limb—something we have not been able to assess experimentally. However, we do not assess the parameter values in detail for the natural storm event since this event could have also included inputs from storm runoff into the stream, which was not measured, while the artificial floods caused remobilization only from the bed. Significant amounts of E. coli can wash into streams with surface runoff water during storm events (Boithias et al., 2021) with in-stream concentrations linked to land use (Bradshaw et al., 2016; Pandey et al., 2018). Surface runoff during

DRUMMOND ET AL. 7 of 10

Conditions

storm events likely explains the gradual decrease in E. coli concentrations during the falling limb as compared to

storm events likely explains the gradual decrease in *E. coli* concentrations during the falling limb as compared to the sharper decrease in model output concentrations (Figure 2). In general, by only including hyporheic exchange flow and release of contaminant microbes from the streambed during stormflow, we were able to represent the variation in concentrations often observed in streams under dynamic flow and advance toward predicting how microbes are transported between the zones (i.e., surface water, hyporheic, and streambed).

4. Conclusions

Our new model framework for fine particle and contaminant microbe transport, hyporheic exchange flow, immorphy bilization, and remobilization during both baseflow and stormflow was able to represent both a series of three artificial floods and a two-peak natural storm event. The model captures the dynamic transport between stream zones with quick exchange into and out of the hyporheic region and slow release from the streambed, so contributing to mechanistic understanding of contaminant microbe accumulation patterns in streams under variable flow and this model framework and differential deposition and resuspension during flashy versus subdued storm hydrogen the stream of the conditions. Natural variation in microbe concentrations during baseflow and stormflow can be represented by this model framework and differential deposition and resuspension during flashy versus subdued storm hydrogen the stream of the hydrogen transport in the falling library to the sharper of the falling library to the stream of the stream of the falling library to the stream of the falling library to the sharper of the falling library to the stream of the stream of the fall of the sharper of the stream of the fall of the sharper zones with quick exchange into and out of the hyporheic region and slow release from the streambed, so contrible uting to mechanistic understanding of contaminant microbe accumulation patterns in streams under variable flow graphs. Future applications of this model framework and differential deposition and resuspension during flashy versus subdued storm hydrog graphs. Future applications of this model to storms in series, accounting for legacy effects from previous storming and the replenishment of microbes in the sediments between events, should further improve characterization of contaminant microbe behavior during both baseflow and stormflow. This should, in turn, assist with assessing waterborne microbial hazards.

Conflict of Interest

The authors declare no conflicts of interest relevant to this study.

Data Availability Statement

The model was implemented in the C++ programming language and is available under an open-source license. **Programming language and is available under an open-source license. **Programming language and is available under an open-source license. **Programming language and is available under an open-source license. **Programming language and is available under an open-source license. **Programming language and is available under an open-source license. **Programming language and is available under an open-source license. **Programming language and is available under an open-source license. **Programming language and is available under an open-source license. **Programming language and is available under an open-source license. **Programming language and is available under an open-source license. **Programming language and is available under an open-source license. **Programming language and is available under an open-source license. **Programming language and is available under an open-source license. **Programming language and is available under an open-source license. **Programming language and is available under an open-source license. **Programming language and is ava

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DRUMMOND ET AL. 9 of 10



10 of 10 DRUMMOND ET AL.